



Selective amplification of ipRGC signals accounts for interictal photophobia in migraine

Harrison McAdams^a, Eric A. Kaiser^b, Aleksandra Igdalova^b, Edda B. Haggerty^b, Brett Cucchiara^b, David H. Brainard^c, and Geoffrey K. Aguirre^{b,1}

^aDepartment of Neuroscience, Perelman School of Medicine, University of Pennsylvania, Philadelphia, PA 19104; ^bDepartment of Neurology, Perelman School of Medicine, University of Pennsylvania, Philadelphia, PA 19104; and ^cDepartment of Psychology, University of Pennsylvania, Philadelphia, PA 19104

Edited by Marlene Behrmann, Carnegie Mellon University, Pittsburgh, PA, and approved June 5, 2020 (received for review April 20, 2020)

Second only to headache, photophobia is the most debilitating symptom reported by people with migraine. While the melanopsin-containing intrinsically photosensitive retinal ganglion cells (ipRGCs) are thought to play a role, how cone and melanopsin signals are integrated in this pathway to produce visual discomfort is poorly understood. We studied 60 people: 20 without headache and 20 each with interictal photophobia from migraine with or without visual aura. Participants viewed pulses of spectral change that selectively targeted melanopsin, the cones, or both and rated the degree of visual discomfort produced by these stimuli while we recorded pupil responses. We examined the data within a model that describes how cone and melanopsin signals are weighted and combined at the level of the retina and how this combined signal is transformed into a rating of discomfort or pupil response. Our results indicate that people with migraine do not differ from headache-free controls in the manner in which melanopsin and cone signals are combined. Instead, people with migraine demonstrate an enhanced response to integrated ipRGC signals for discomfort. This effect of migraine is selective for ratings of visual discomfort, in that an enhancement of pupil responses was not seen in the migraine group, nor were group differences found in surveys of other behaviors putatively linked to ipRGC function (chronotype, seasonal sensitivity, presence of a photic sneeze reflex). By revealing a dissociation in the amplification of discomfort vs. pupil response, our findings suggest a postretinal alteration in processing of ipRGC signals for photophobia in migraine.

ipRGCs | melanopsin | migraine | photophobia

People find bright light uncomfortable and sometimes even painful. This experience of light-induced discomfort is exacerbated in numerous clinical conditions and can be debilitating (1). We refer here to discomfort from light as photophobia, which is typically manifest as a somatic sensation localized to the eyes or head (2). A common cause of photophobia is migraine (3). Photophobia is reported by 80 to 90% of individuals during a migraine attack (4–6), and 50% of individuals report it as their most burdensome symptom (7). Even between headaches, people with migraine have a lowered threshold for pain from light as compared with headache-free controls (Haf) (8–11).

The signals that ultimately result in photophobia presumably begin with photoreceptors in the eye. Under daylight conditions, the cone photoreceptors capture photons and relay signals via retinal ganglion cells to thalamic and brainstem targets (Fig. 1A). A subset of retinal ganglion cells expresses the photopigment melanopsin (12). These intrinsically photosensitive retinal ganglion cells (ipRGCs) are capable of responding to light without synaptic input (13). There is evidence from rodent studies that ipRGCs project to the somatosensory thalamus, where they innervate neurons that are also sensitive to dural stimulation carried by trigeminal afferents (14) (Fig. 1A). This finding offers a neural mechanism by which light stimulation creates somatic discomfort. The ipRGCs contribute to several other “reflexive” functions of vision as well, most notably photoentrainment of the circadian rhythm (15, 16) and control of pupil size (17–19).

The ipRGCs may play a role in human photophobia. People who have migraine and are also blind from inherited rod–cone degeneration experience photophobia during a headache (14), implicating spared ipRGCs as the source of this sensation. In people without visual impairment, Stringham and colleagues (20) found that shorter wavelengths of light (closer to the peak spectral sensitivity of the melanopsin photopigment) tend to produce greater discomfort in healthy observers (20). Studies that use narrow-band light stimuli, however, are limited in their ability to probe the specific contribution of melanopsin to photophobia in the intact visual system. This is due to the considerable overlap of the cone and melanopsin spectral sensitivity functions (Fig. 1B). Moreover, some classes of ipRGCs also receive input from the cones (13, 21–23). As a consequence, photophobia may result from both melanopsin and cone signals after their integration within ipRGCs. It is unknown how these photoreceptor classes are weighted and combined to produce photophobia and how this process might be altered in migraine.

In previous work, we have shown that carefully tailored modulations of the spectral content of light may be used to selectively target melanopsin or the cones (24, 25). Here, we examine the contribution of cone and melanopsin signals to visual discomfort, and to pupil responses, in people who have migraine

Significance

The melanopsin-containing intrinsically photosensitive retinal ganglion cells (ipRGCs) may contribute to photophobia in migraine. We measured visual discomfort and pupil responses to cone and melanopsin stimulation—the photoreceptor inputs to the ipRGCs—in people with and without migraine. We find that people with migraine do not differ from those without headaches in how cone and melanopsin signals are weighted and combined to produce visual discomfort. Instead, migraine is associated with amplification of ipRGC signals for discomfort. This effect of migraine upon ipRGC signals is limited to photophobia, as we did not find an enhancement of pupil responses or a change in other behaviors linked to ipRGC function. Our findings suggest a postretinal amplification of ipRGC signals for photophobia in migraine.

Author contributions: H.M., E.A.K., B.C., D.H.B., and G.K.A. designed research; H.M., A.I., and E.B.H. performed research; H.M., E.B.H., and G.K.A. analyzed data; and H.M., E.A.K., and G.K.A. wrote the paper.

Competing interest statement: E.A.K. has received royalties from a patent shared with Alder Biopharmaceuticals. The remaining authors declare that they have no relevant financial interests that relate to the research described in this paper.

This article is a PNAS Direct Submission.

Published under the PNAS license.

Data deposition: The raw data and analysis code are available through GitHub (<https://github.com/gkaguirrelab/melSquintAnalysis>).

¹To whom correspondence may be addressed. Email: aguirreg@upenn.edu.

This article contains supporting information online at <https://www.pnas.org/lookup/suppl/doi:10.1073/pnas.2007402117/-DCSupplemental>.

First published July 6, 2020.

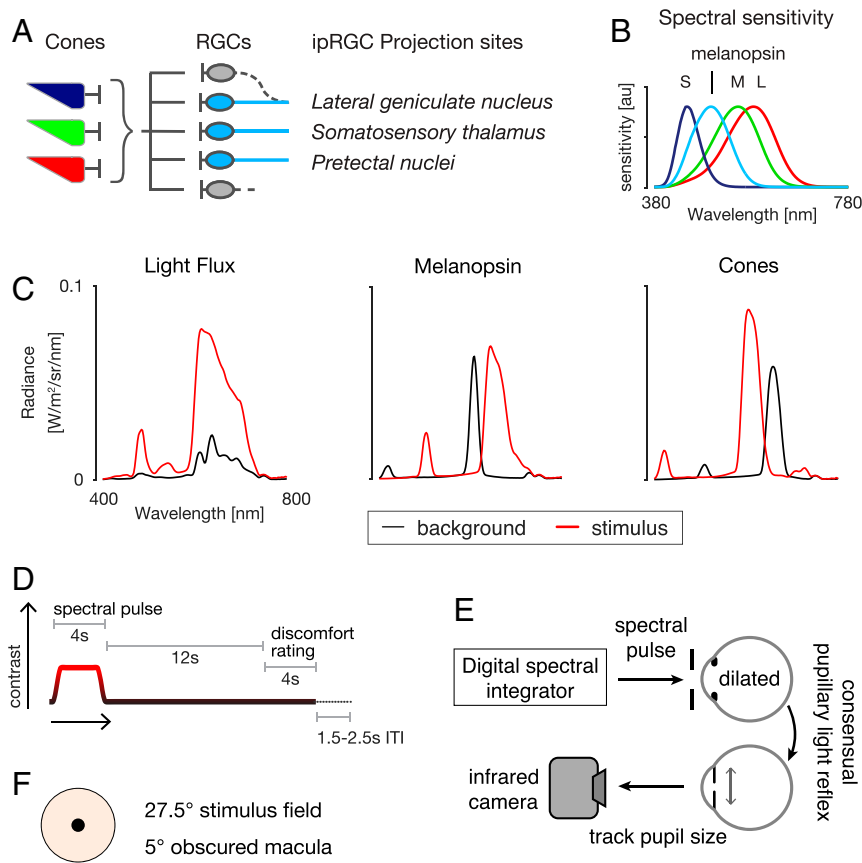


Fig. 1. Experiment overview. (A) There are several classes of melanopsin-containing ipRGCs, which vary in their central projections, function, and extent to which they receive input from cones. The ipRGCs project to the somatosensory thalamus and the lateral geniculate nucleus, where their signals may contribute to light sensitivity. Other ipRGCs project to the pretectal nuclei to control the size of the pupil. Not shown are numerous, additional subcortical projection targets of the ipRGCs (e.g., the suprachiasmatic nucleus). RGC, retinal ganglion cell. (B) The spectral sensitivity functions of the relevant photoreceptors under daylight conditions. S, M, and L refer to the short-, medium-, and long-wavelength sensitive cones, respectively. (C) Shown are pairs of spectra (background: black; stimulus: red) that differ in excitation for the targeted photoreceptors. In *Left*, *Center*, and *Right*, the stimuli produce equal contrast on the cones and melanopsin (termed light flux), contrast only on melanopsin, and equal contrast across all three classes of cones but no contrast on melanopsin, respectively. (D) Each trial featured a 4-s period during which the stimulus transitioned from the background to the stimulation spectrum and back. Twelve seconds after stimulus offset, the subject provided a discomfort rating. There was an intertrial interval that varied between 1.5 and 2.5 s. (E) The light from a digital spectral integrator was presented to the pharmacologically dilated right eye of the subject through an artificial pupil. The consensual pupillary light response of left eye was recorded with an infrared camera. (F) The stimulus spectra were presented in an eyepiece with a 27.5°-diameter field, with the central 5° obscured to minimize macular stimulation.

with interictal photophobia. Participants reported the discomfort they experienced from viewing pulses of light that selectively targeted melanopsin, the cones, or their combination (Fig. 1 C and D). Pupillometry in response to these pulses was also obtained (Fig. 1E). We recruited 20 participants in each of three groups: migraine with visual aura (MwA), migraine without aura (MwoA), and HAF. All of the participants with migraine endorsed interictal sensitivity to light. Our findings demonstrate that both melanopsin and cone stimulation in isolation produce visual discomfort. By examining the effect of separate and simultaneous stimulation of melanopsin and the cones, we quantified how these photoreceptor signals are weighted and combined to produce visual discomfort and pupil responses. We find that the enhanced interictal light sensitivity observed in migraine is well described as an amplification of photoreceptor signals after their combination. We further demonstrate that pupil responses are governed by different combination parameters and do not demonstrate amplification in migraine. These results indicate that interictal photophobia in migraine is a selective amplification of a subset of ipRGC outputs, most plausibly at a postretinal locus.

Results

Participant Demographic and Clinical Characteristics. We studied 20 people from each of three groups: MwA, MwoA, and HAF. The three groups (Table 1) were well matched in age ($F[2,57] = 0.2$, $P = 0.820$) but differed in gender distribution ($F[2,57] = 3.3$, $P = 0.0439$), with fewer women in the control group. The greater proportion of women in the migraine groups is consistent with migraine epidemiology (26). Headache frequency was similar in the two migraine groups with 12 (± 10) and 13 (± 9) days with headache reported within a 90-d period by MwA and MwoA subjects, respectively (approximately four headache days per month), consistent with a classification of episodic (as opposed to chronic) migraine (27). Acetaminophen and nonsteroidal anti-inflammatory drug (NSAID) use (for any indication) were similar across all three groups. Triptan use was reported by five MwA participants and one MwoA participant. Similarly, combined aspirin/acetaminophen/caffeine (Excedrin) use was reported by six MwA participants and one MwoA participant. Preventive medication use (e.g., tricyclics, beta-blockers, etc.) was reported by one MwA participant and three MwoA participants. We quantified headache disease burden using the Migraine Disability Assessment Test (MIDAS) (28) and Headache

Table 1. Subject demographic and clinical characteristics

Group	No. of women	Age (y)	Headache days/3 mo	Disability		Medication use last 3 mo				
				MIDAS	HIT-6	NSAID	Actmnphn	Excedrin	Triptan	Preventive
Controls	13/20	31 (5)	1.3 (1.4)	0.5 (0.8)	40.7 (4.1)	14	1	0	0	0
MwA	19/20	31 (4)	13.1 (8.9)	18.6 (15.3)	60.6 (8.0)	16	5	6	5	1
MwoA	17/20	30 (4)	11.7 (9.7)	16.0 (13.7)	60.0 (8.8)	17	2	1	1	3

Participants were asked to report the number of headaches they had experienced over the prior 3 mo. The MIDAS (28) and the HIT-6 (29) measure headache disability. Where appropriate, the mean value (and SD) across subjects is reported. Medication use is summarized within five categories: actmnphn, acetaminophen for any indication; Excedrin, use of any one of multiple formulations that combine an NSAID with acetaminophen to treat headache; NSAID, NSAID for any indication; preventive, any one of several classes of medications used to decrease migraine frequency (e.g., tricyclic antidepressants, beta-blockers); triptan, any tryptamine-based drug used to abort a migraine.

Impact Test (HIT-6) (29) surveys. The migraine groups unsurprisingly had higher scores on both instruments relative to HAF (MIDAS: $F[2,57] = 13.65$, $P = 1.43e-5$; HIT-6: $F[2,57] = 48.82$, $P = 4.43e-13$). The two migraine groups did not differ in disease impact (MIDAS: $t = 1.00$, $P = 0.76$; HIT-6: $t = 0.40$, $P = 0.96$). The distribution of these values suggests moderate disability from migraine in both groups.

Participants with Migraine Have Interictal Photophobia but Do Not Differ from Controls in Surveys of Circadian and Seasonal Behavior.

The Visual Discomfort Scale (VDS) measures symptoms of discomfort from reading, patterns, and light on a 0 to 69 scale (30). We required our control participants to have a low score on this instrument (less than or equal to seven) but did not impose a requirement for migraineurs. Symptoms of visual discomfort were correspondingly greater in the migraine population as compared with the controls (Table 2) ($F[2,57] = 15.23$, $P = 5.02e-6$). Participants also completed the Photosensitivity Assessment Questionnaire (PAQ), which measures light-avoiding (“photophobia”) and light-seeking (“photophilia”) behavior on a zero to eight scale (31). Migraine participants again reported greater light avoidance as compared with controls (Table 2) ($F[2,57] = 10.95$, $P = 9.44e-5$), although there was no difference in reported light-seeking behavior (Table 2) ($F[2,57] = 0.75$, $P = 0.448$).

As we are interested in how migraine and photophobia may relate to ipRGC function, we examined if our participant groups differed in other functions thought to be mediated by ipRGCs. In the rodent, multiple classes of ipRGCs have been identified that differ in their subcortical projections and in their functional properties. Projections of the ipRGCs to the suprachiasmatic nucleus are thought to control circadian photoentrainment (15). As variation in this function is speculated to relate to sleep alterations and seasonal affective disorder, we gathered information about the sleep habits and seasonal preferences of our participants (Table 2). The Morningness-Eveningness Questionnaire (32) characterizes chronotype on a scale of 16 to 86, with the extremes corresponding to evening and morning preference, respectively. The median scores for the three groups all

were in the midrange (~50) and were not significantly different ($F[2,57] = 0.54$, $P = 0.586$). The Seasonal Pattern Assessment Questionnaire (33) provides a Global Seasonality Score, which assesses on a 0 to 24 scale the degree to which mood and physiology vary across seasons; a score of 16 or higher is typical in patients with seasonal affective disorder. The central tendency of our participants (a score of approximately seven) indicates a mild degree of seasonal sensitivity, and this did not differ between the groups ($F[2,57] = 2.19$, $P = 0.121$). Finally, the photic sneeze reflex has been hypothesized to be related to ipRGC function (34), as the somatic “prickling” sensation of the nose in response to a bright light is suggestive of a retinotrigeminal interaction. We asked our participants if they experience this phenomenon and did not find any difference between groups in the proportion of participants (15 to 20%) who have this experience ($F[2,57] = 0.04$, $P = 0.962$).

Overall, apart from photophobia, our studied populations were well matched in behaviors hypothesized to be related to ipRGC function.

Melanopsin and Cone Contrast Produce Mild Discomfort in Control Participants.

Our participants rated the degree of discomfort they experienced while viewing pulses of spectral change that targeted melanopsin, the cones, or combined stimulation of both sets of photoreceptors (termed light flux). The stimuli were designed to increase excitation in the targeted photoreceptor(s) by 100, 200, or 400%. Participants rated the amount of discomfort produced by each type of light pulse on a 0 (none) to 10 (extreme) scale.

The light flux stimulus combines melanopsin and cone stimulation. In the HAF participants, light flux pulses evoked mild discomfort, increasing with contrast, reaching a mean discomfort rating of 3.15 of 10 for 400% contrast (Fig. 2, *Top Left*). To determine whether this discomfort was a consequence of melanopsin or cone-based signaling, we examined the discomfort ratings in response to stimuli designed to target these photoreceptor classes in isolation. Discomfort ratings to both melanopsin (Fig. 2, *Middle Left*) and cone-directed stimuli (Fig. 2, *Bottom Left*) also increased with contrast but only with mild discomfort at 400% (Fig. 2, *Middle Left* [mean rating of 2.18 for melanopsin]

Table 2. Surveys of behaviors that may be related to ipRGC function

Group	VDS	PAQ–photophobia	PAQ–photophilia	SPAQ	Morningness-Eveningness	Photic sneeze reflex
Controls	3.45 (1.76)	0.15 (0.19)	0.71 (0.21)	6.10 (3.81)	51.55 (10.28)	4
MwA	16.55 (10.00)	0.46 (0.28)	0.65 (0.22)	8.90 (5.53)	48.80 (10.53)	3
MwoA	13.15 (8.88)	0.51 (0.31)	0.63 (0.23)	8.65 (4.55)	48.75 (8.32)	3

The VDS measures reported light sensitivity across several domains of visual function (30). The PAQ measures reported photophobia and photophilia behaviors (31). The Seasonal Pattern Assessment Questionnaire (SPAQ) measures the reported degree to which mood and behavior vary over course of a year (33), and the Morningness-Eveningness Questionnaire provides a “chronotype” score (32). Values are the mean (and SD) across subjects within each group. Finally, we asked subjects if they “tend to sneeze when [they] step out of a dark room into bright sunlight” and report here the number of subjects in each group who responded “yes.”

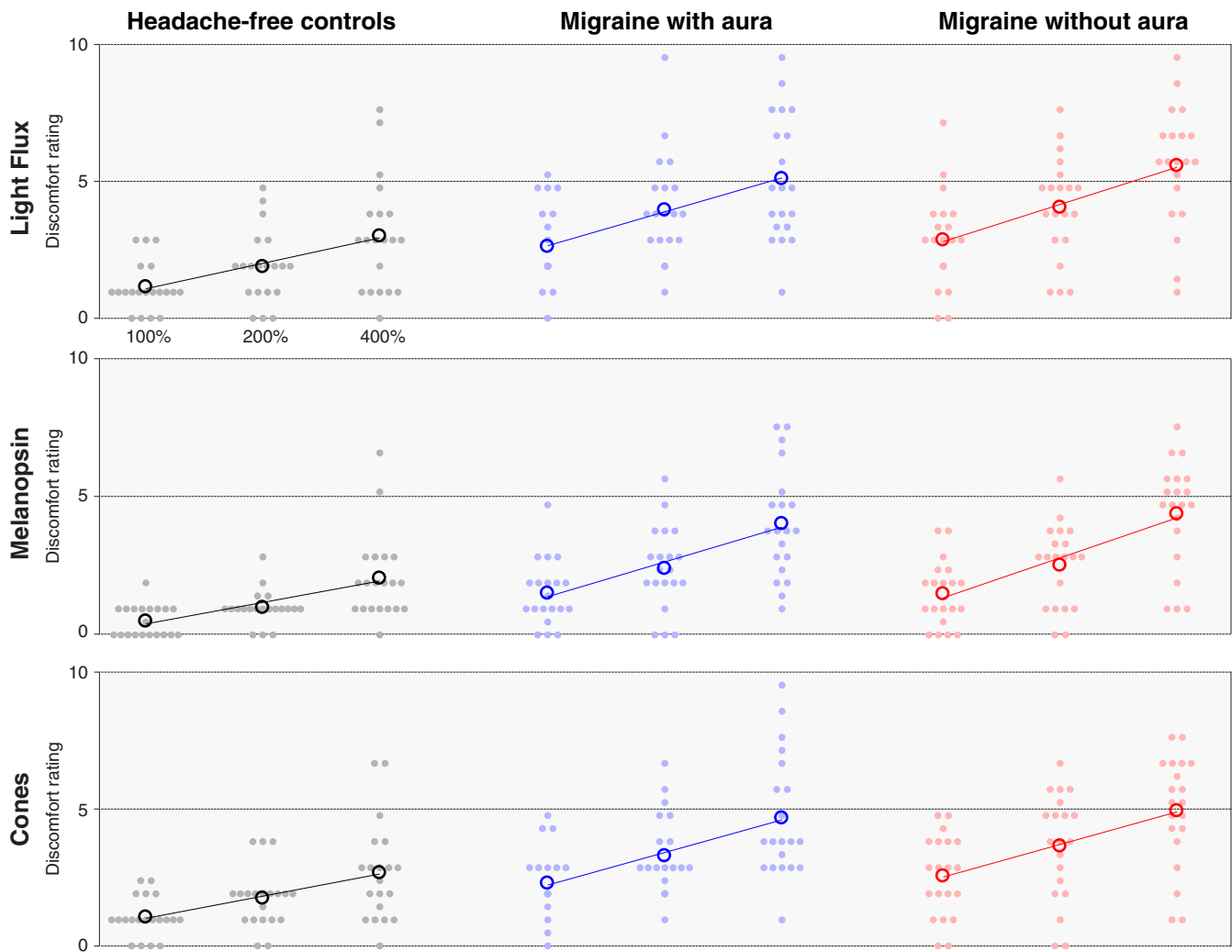


Fig. 2. Discomfort ratings by stimulus and group. Each row presents the discomfort ratings elicited by stimuli that targeted a particular combination of photoreceptors, and each column contains the data from each individual group ($n = 20$ participants per group). The stimuli were presented at three different contrast levels (100, 200, and 400%), and these (log-spaced) values define the x axis of each subplot. The median (across trial) discomfort rating for a given stimulus and contrast is shown for each participant (filled circles), as is the mean rating across participants (open circles). The best-fit line to the mean discomfort rating across participants as a function of log contrast is shown in each subplot.

and *Bottom Left* [2.80 for cones]). This result suggests that both cone and melanopsin signals contribute to light-induced discomfort. For all stimuli, we further observed that logarithmic changes in stimulus contrast produced linear changes in mean rated discomfort, as illustrated by the good agreement between the fit lines and the data (Fig. 2).

Cone and Melanopsin Signals Contribute to Interictal Photophobia in Migraine. We next asked if people with photophobic migraine would experience greater discomfort in response to our stimuli and if so, whether the enhanced discomfort signal is attributable to the cones, melanopsin, or both. Both migraine groups showed increased discomfort in response to the combined light flux stimuli at all contrast levels (Fig. 2, *Top Center* and *Top Right*: at 400% contrast, mean of 5.35 for MwA and 5.85 for MwoA vs. 3.15 for controls). The mean rating across participants was also increased in both migraine groups in response to melanopsin-directed stimulation (Fig. 2, *Middle*: at 400% contrast, mean of 4.28 for MwA and 4.65 for MwoA vs. 2.18 for controls) and cone-directed stimulation (Fig. 2, *Bottom*: at 400% contrast, mean of 4.90 for MwA and 5.18 for MwoA vs. 2.80 for controls). Both

migraine groups also showed a linear relationship between log-spaced contrast and mean discomfort ratings for all stimulus types, which is again illustrated by the fit lines (Fig. 2). A mixed effects ANOVA confirmed that discomfort ratings were higher for all stimuli in the migraine groups as compared with the control group (*SI Appendix, Table S2*).

There was a higher proportion of women in the migraine groups as compared with the control group. We considered if this unequal distribution of gender could account for the differences in discomfort ratings between the groups. The mean discomfort rating reported by female control participants (across all stimuli) was not higher than the ratings provided by male participants (mean rating men: 1.79; women: 1.74), indicating that differing gender ratios do not account for the increased discomfort in the migraine groups.

Discomfort Ratings Are Well Fit by a Two-Stage, Nonlinear, Log-Linear Model. We observe that mean discomfort ratings for all stimuli are well described as a linear function of log-scaled stimulus contrast, consistent with the Weber–Fechner law of perception. It is also apparent that a light flux stimulus, which combines

melanopsin and cone contrast, evokes less discomfort than would be predicted given the discomfort produced by each stimulus component alone. These properties of the data may be explained by nonlinear combination of melanopsin and cone signals prior to the stage at which photoreceptor signals are interpreted as discomfort.

We examined these impressions within the context of a quantitative, two-stage model governed by four parameters. The first stage is based upon psychophysical measures of combined stimulus dimensions (35, 36), and the second is on the Weber–Fechner law. The model provides a discomfort rating for stimuli with arbitrary combinations of melanopsin and cone contrast.

The first stage of the model (Fig. 3 A, Left) considers the combination of melanopsin and cone signals within ipRGCs. The inputs to this stage are the contrasts on the melanopsin and cone photoreceptors created by a stimulus. A light flux stimulus of (for example) 200% contrast has the property of providing 200% contrast input on both of these photoreceptor classes. A scaling factor (α) adjusts the relative potency of melanopsin contrast as compared with cone contrast. The two contrast types are then combined using a Minkowski distance metric with exponent β .

This integrated “ipRGC contrast” is log transformed and then passed to the second stage of the model (Fig. 3 A, Right), which transforms input into a discomfort rating under the control of a slope and offset parameter (which is the intercept transformed to describe the modeled response to 200% contrast).

We fit this model to the discomfort ratings across trials for all stimuli and participants within a particular group using bootstrap resampling across participants to characterize the variability of the model parameters. Fitting was performed separately for the data from each group (Fig. 3 B). The model performed equally well for each group in accounting for the mean (across-participant) discomfort ratings across stimuli (model $R^2 \pm$ SEM: HAF: 0.95 ± 0.03 ; MwA: 0.96 ± 0.03 ; MwoA: 0.97 ± 0.01).

Migraine Groups Differ from HAF in the Response to Integrated Melanopsin and Cone Signals. We examined the fitted parameters of the model and compared these values across groups (Fig. 3 B). The discomfort data from all three groups are best fit by first scaling (α) the influence of melanopsin contrast by $\sim 60\%$. The scaled melanopsin and cone contrast are then combined with a subadditive Minkowski exponent (β) of ~ 1.75 , intermediate between

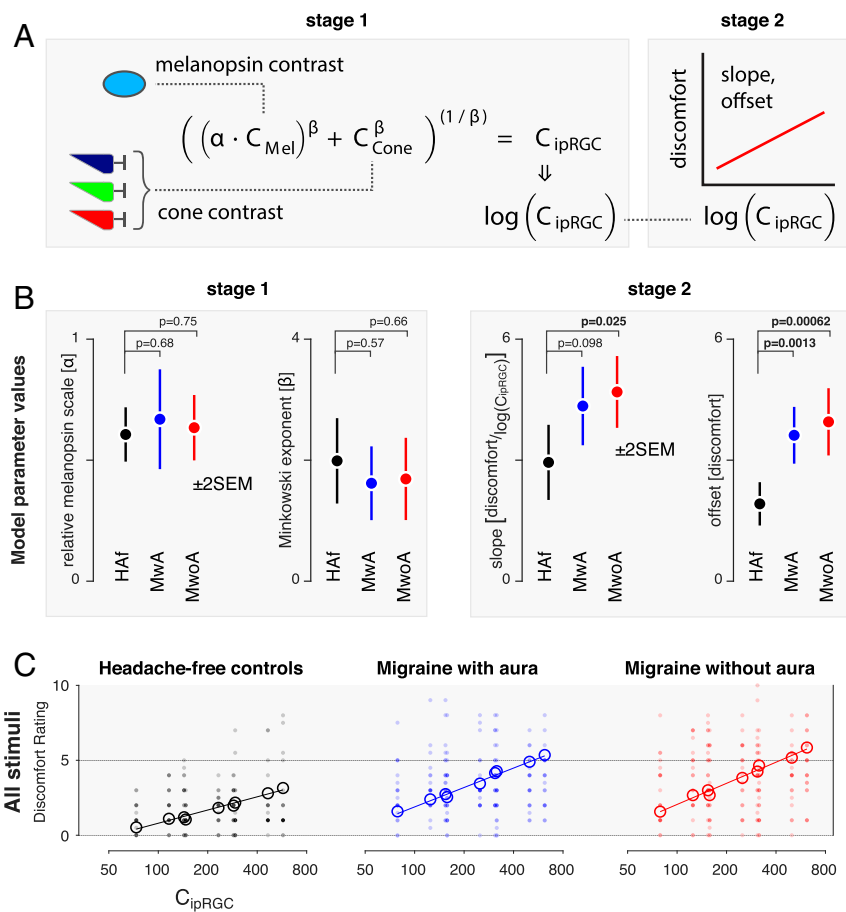


Fig. 3. A two-stage model of discomfort ratings. We developed a two-stage model that describes discomfort ratings as a function of melanopsin and cone stimulation. (A) In the first stage, (Left) melanopsin contrast (C_{Mel}) is weighted by a scaling factor (α) and then combined with cone contrast (C_{Cone}) under the control of the Minkowski exponent (β). The output of this stage is ipRGC contrast, which is log transformed and passed to the second stage (Right). Here, the signal undergoes an affine transform to produce a discomfort rating, under the control of a slope and offset parameter (the latter being expressed as the modeled discomfort rating at 200% ipRGC contrast). (B) The model was fit to the discomfort data from each group, yielding estimates of the model parameters (± 2 SEM obtained via bootstrapping). The P value associated with a two-tailed t value, taken with respect to the pooled SEs, is presented for the comparison of each of the migraine groups with the control group for each parameter ($n = 20$ participants per group). (C) Stage 1 of the model transforms the stimuli used in the experiment to common units of ipRGC contrast. Each plot presents the discomfort ratings (individual participants in filled circles, group means in open circles) in terms of ipRGC contrast, with the parameters at stage 1 forced to be the same across groups. The nine open circles correspond to the nine stimuli used in the experiment (three contrast levels each of melanopsin, cone, and light flux stimulation). The fit of the second stage of the model (which can vary across groups) provides the fit line.

simple additivity ($\beta = 1$) and a Euclidean distance metric ($\beta = 2$). We find that these parameter values do not significantly differ between the three groups (Fig. 3 *B, Left*). Therefore, we do not find that people with photophobic migraine differ from HAF in the manner in which melanopsin and cone signals are combined at this initial stage.

The second pair of parameters converts log-transformed ipRGC contrast into discomfort ratings. Here, substantial differences between the migraine and control groups were found. The MwoA group had a greater slope and a higher offset of discomfort rating, and the Mwa group had a higher offset, for a given amount of ipRGC contrast (Fig. 3 *B, Right*). The migraine groups reported discomfort that was roughly twice as great overall and had a slope that was 50% steeper as compared with controls for the increase in discomfort with ipRGC contrast.

Based upon these results, we refit the model, forcing the stage 1 parameters to be the same across the three groups but allowing the stage 2 parameters to vary. The output of stage 1 allows us to describe all of the stimuli used in the experiment in terms of a single value of ipRGC contrast. Fig. 3C replots the discomfort data for all participants and all stimuli from each group in terms of the stage 1 value of ipRGC contrast. The stage 2 model fits differ for each group and are used to generate the solid lines on the plots. Open circles mark the mean, across-participant discomfort ratings for each of the nine stimulus types. There is good agreement between the model fit and the across-participant mean discomfort. Forcing the stage 1 parameters to be the same across groups had minimal impact upon the fit of the model to the data (model $R^2 \pm$ SEM: HAF: 0.95 ± 0.03 ; Mwa: 0.96 ± 0.02 ; MwoA: 0.97 ± 0.01), supporting the claim that the stage 1 model parameters do not meaningfully differ between the groups.

Overall, these findings indicate that people with migraine with interictal photophobia do not differ from controls in the manner in which cone and melanopsin signals are scaled relative to each other and combined but experience greater discomfort from this integrated signal.

Migraine Groups Do Not Have Enhanced Pupil Responses, Indicating a Selective Enhancement of ipRGC Discomfort Signals. We considered the possibility that people with migraine have a general amplification of ipRGC signals at the level of the retina, of which visual discomfort is one aspect. If so, then we might expect an amplification of pupil responses to be seen in this population as well. To test this idea, we compared pupil constriction in the migraine groups with that observed in the HAF participants.

Fig. 4A presents the mean, across-participant pupil responses observed in each of the three groups to the stimuli used in the experiment. The temporal profile of the pupil response to stimuli that target melanopsin, the cones, or their combination is in good agreement with prior reports (25). There is also a clear increase in the amplitude of the pupil constriction produced by stimuli with increasing (100, 200, 400%) contrast.

The responses obtained from each studied group are close to overlapping in the plots for each combination of stimulus direction and contrast. We did not observe a greater amplitude of pupil response in the migraine groups as compared with the controls. Indeed, if anything, the pupil response in the migraine groups (particularly MwoA) is slightly attenuated compared with that of the HAF. We quantified the pupil response for each participant by measuring the mean percent change in pupil area following stimulus onset (*SI Appendix, Fig. S1*). Similar to what was observed for visual discomfort ratings, the relationship between pupil response and stimulus contrast is well described as log linear.

We next examined how cone and melanopsin signals are combined to produce the overall amplitude of pupil constriction, using the same two-stage model that we developed for the discomfort ratings (Fig. 4B). The model fit the data from the three

groups well (model $R^2 \pm$ SEM: HAF: 0.95 ± 0.03 ; Mwa: 0.98 ± 0.01 ; MwoA: 0.94 ± 0.02). We found no significant differences between the groups in the parameters of the model for pupil response. Therefore, we refit the model to the pupil data, forcing all parameters to be the same across the groups (Fig. 4C). The agreement between the data and the model was quite good, despite requiring that all three groups be described using the same model parameters (model $R^2 \pm$ SEM: HAF: 0.96 ± 0.01 ; Mwa: 0.95 ± 0.001 ; MwoA: 0.91 ± 0.06).

The stage 1 parameters in control of the pupil describe a scaling factor for melanopsin (α) of $\sim 40\%$, which is somewhat less than the influence that melanopsin has upon discomfort ($\sim 60\%$). The Minkowski exponent for the combination of melanopsin and cone signals in the pupil response is ~ 0.8 , compared with its value of ~ 1.75 for the discomfort ratings. The value of ~ 0.8 indicates a combination rule for cone and melanopsin signals that is reasonably close to linear, consistent with prior observations of the additivity of cone and melanopsin signals in the pupil response (24, 37). The fact that the stage 1 parameters differ between the model fits to the two measures indicates that discomfort and pupil control are mediated by mechanisms that combine signals from melanopsin and the cones in different ways. A possible neural basis for these mechanisms would be distinct classes of ipRGCs.

Separately from the matter of how signals from melanopsin and the cones are combined across the two measures, the fact that the stage 2 parameters differ between controls and people with migraine for the production of discomfort but not for pupil constriction argues against the idea that a common, single amplification of retinal signals mediates increased interictal photophobia in migraine.

Discussion

Our study indicates that the enhanced, interictal light sensitivity experienced by people with migraine is due to a selective amplification of response to a subset of ipRGC signals. Photophobia in migraine is not the result of an omnibus change in cone or melanopsin signals per se but instead, a change in the response to these photoreceptor inputs after they have been weighted and combined. Moreover, this enhanced response is specific for discomfort signals in that it is not observed for ipRGC outputs that control other reflexive responses to light, in particular pupil constriction.

Distinct ipRGC Classes. Studies in rodents (38–40), primates (13, 41–43), and in the postmortem human eye (44, 45) have demonstrated the existence of multiple classes and subclasses of ipRGCs, which differ in their photoreceptor inputs, signaling kinetics, and central projections. Control of circadian photoentrainment and the pupil response, for example, is attributable to distinct subsets of ipRGCs in rodents (38, 46).

We examined how melanopsin and cone inputs contribute separately and in combination to visual discomfort and to the pupil response within the context of a quantitative model. The first stage of our model estimates how melanopsin signals are weighted relative to cone signals and the metric with which melanopsin and cone signals are combined. We did not find a difference at this stage between people with or without migraine. We did find, however, that the model parameters differ substantially when measured for the pupil response and for ratings of visual discomfort. A plausible explanation for these differences in photoreceptor combination is that different classes of ipRGCs contribute to visual discomfort and pupil responses in the human.

In the current study, we find that melanopsin and cone signals are combined approximately additively in control of the pupil, consistent with prior reports (24, 37). Melanopsin contrast was 40% as effective as cone contrast in modulating the pupil for

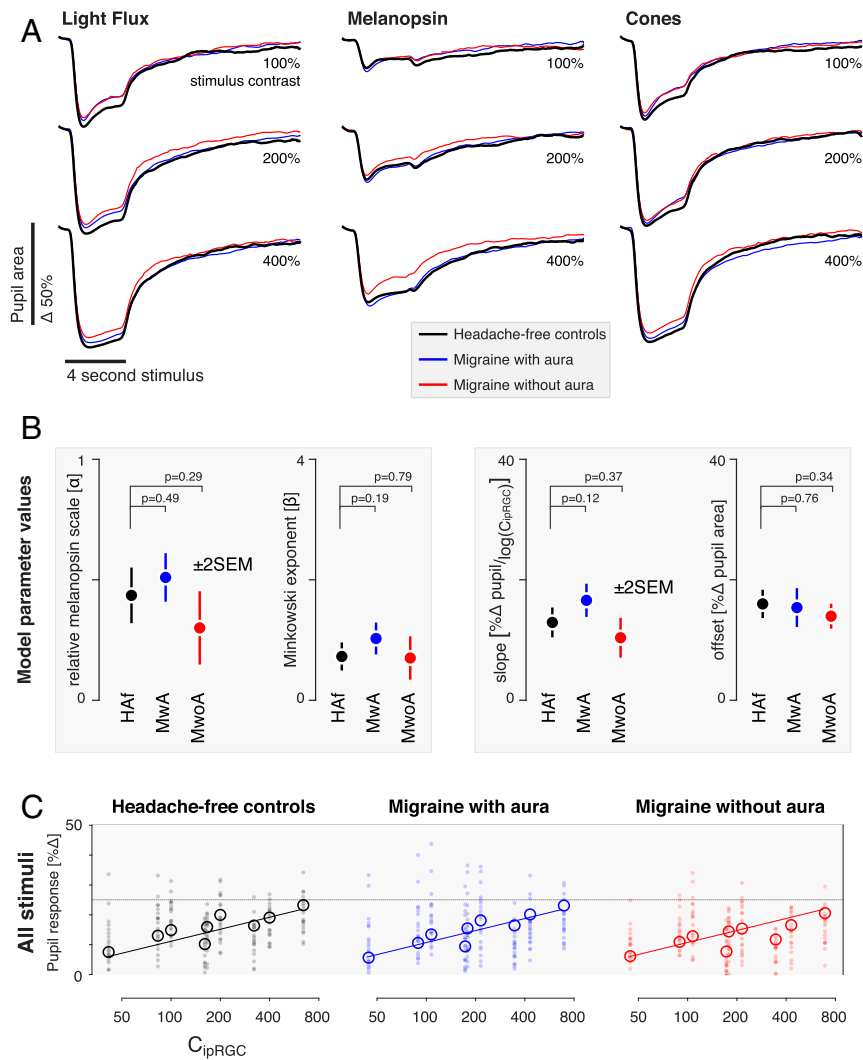


Fig. 4. Pupil response by stimulus and group. (A) The average pupil response across participants within each group ($n = 20$ participants per group) is shown for each stimulus type (columns) at each contrast level (rows). The responses from the three groups for each stimulus type are superimposed. (B) We summarized the pupil responses by taking the mean of the percent change in amplitude of the pupil area across the recording period. These data were then fit with the two-stage model (Fig. 3). No significant differences between the groups in the parameter estimates were found (± 2 SEM obtained via bootstrapping), although both the relative melanopsin scaling and Minkowski exponent values are smaller for pupil responses than was observed for discomfort ratings. (C) As no significant differences between groups were found for the parameters, we refit our model to the data forcing all parameters to be the same across groups. The plots report individual (filled circles) and mean (open circles) pupil response as a function of modeled ipRGC contrast.

these pulsed stimuli, as compared with a prior report of an overall 26% effectiveness of melanopsin relative to modulations that placed equal contrast on long- and medium-wavelength sensitive cones for driving pupil responses with sinusoidal modulations of contrast at low and high temporal frequencies (24). We note that our index of pupil change here was across the entire time course of evoked response. While it is likely that the relative contribution of melanopsin to the amplitude of pupil constriction varies as a function of time following stimulus onset (19, 25, 37, 47, 48), such a dissection of the components of the pupil response is beyond the scope of the current report.

In contrast to the pupil response, melanopsin and cone signals exhibit a nearly Euclidean combination metric in our measure of discomfort, and we find that the influence of melanopsin signals (relative to the cones) is ~ 1.5 times greater in producing visual discomfort as compared with pupil responses. A Euclidean combination metric is a feature of stimulus dimensions that produce a single, integrated percept (35), suggesting that cone

and melanopsin signals are combined into a unitary experience of discomfort.

We have previously found that observers describe targeted melanopsin stimulation as “uncomfortable brightness” (49). It may be the case that the “brightness” and “discomfort” percepts, while each integrating cone and melanopsin signals, reflect the action of distinct retinal ganglion cell populations. Our present data, however, do not directly address such a dissociation.

Several studies have demonstrated that melanopsin contrast contributes to a sensation of brightness (50–53). The melanopic component of brightness is presumably combined with the postreceptoral luminance channel that is derived from the sum of L and M cone excitations and carried by the classical (nonmelanopsin-containing) retinal ganglion cells. Yamakawa and colleagues (51) measured the perceptual brightness of lights that varied in melanopic and luminance content. A roughly additive effect of luminance and melanopsin content upon brightness is present in their data, although the form of the response

departed from linear. The interpretation of these measurements is complicated, however, as the observers did not undergo pharmacologic dilation of the pupil, causing retinal irradiance to vary systematically with the stimulus.

Zele and colleagues (53) also examined how cone and melanopsin signals combine in the perception of brightness. Their work shows a log-linear relationship between isolated melanopsin and cone stimulus intensity and brightness. However, when presented in combination, they report two contribution components of cones to brightness, one of which is negative and may imply an adaptation process.

Selective Amplification. The ipRGCs are known to manifest linear changes in firing rates with logarithmic changes in retinal irradiance (54). In our measurements, we find that ratings of visual discomfort and the amplitude of evoked pupil response vary linearly with log changes in stimulus contrast, consistent with an output system that receives these log-transformed signals from the ipRGCs.

While participant groups did not differ in the manner in which cone and melanopsin signals were combined, we find that people with episodic migraine with interictal photophobia have an enhanced response to this integrated signal in their ratings of visual discomfort. This amplification is similar in migraine with or without visual aura.

Importantly, we did not find evidence of a general amplification of ipRGC signals in migraine. The ipRGCs are the dominant, and perhaps exclusive, route for photoreceptor signals influencing the light-evoked pupil response via the pretectal nuclei (55, 56). If migraine is accompanied by a general amplification of ipRGC signals, then an enhanced light-evoked pupil response in this population might be predicted. Instead, we find that the amplitude of evoked pupil responses is not increased in people with migraine in response to stimulation of melanopsin, the cones, or their combination. Indeed, the trend in the data was toward smaller evoked pupil responses in migraine, especially in MwoA. Prior studies of pupil response in migraine have obtained varying results. Prior studies have not found migraine group differences in the amplitude of pupil constriction or steady-state pupil size (57–60), although more subtle changes in pupil dynamics have been reported (58, 61, 62). Our study differs from many prior reports in that we measured open-loop, consensual pupil responses by combining pharmacologic dilation with an artificial pupil, thus controlling retinal irradiance across the studied groups.

We also surveyed our participants regarding other behaviors that may be related to ipRGC function. A general alteration in ipRGC function in people with migraine might be predicted to be manifest in these measures as well. The ipRGCs have been implicated in circadian photoentrainment (15), seasonal variation in mood and physiology (63–65), and in the photic sneeze reflex. Our participants with migraine did not differ from HAF in these behaviors, again suggesting that the amplification of ipRGC signals in migraine is specific to visual discomfort.

The Neural Locus of Amplification. While no specific ipRGC subtype has been identified as carrying the signal for visual discomfort, various lines of evidence implicate ipRGCs distinct from the M1 subtype (66–68). The ipRGCs coinnervate neurons within the posterior thalamus of the rodent that also receive trigeminal afferents. These thalamic neurons then project onward to both somatosensory and visual cortices. Classes of ipRGCs also project to the lateral geniculate nucleus (13, 54) and are capable of modulating visual cortex responses (49). Our findings of amplified discomfort to visual stimulation in people with migraine could reflect alteration of signals derived from the ipRGCs at any one of these sites.

A physiologic hallmark of migraine is alteration in the excitability of cortex, as manifest both in the phenomenon of cortical spreading depression of aura and a tendency toward enhanced responses to sensory stimulation as compared with HAF (69). Enhanced cortical responses to sensory stimulation have been observed in migraine with (69) and possibly without (70) aura, and for multiple sensory modalities. A natural locus, therefore, for the amplification of ipRGC signals for visual discomfort is at cortical sites. This could take place within primary visual or somatosensory cortex or further downstream at the point where multimodal signals are integrated into a report of discomfort.

An ipRGC signal of visual discomfort might also be amplified at the level of the thalamus. Altered thalamic gating has been proposed as a mechanism for altered sensory perception in migraine, including photophobia (3, 71). Enhanced signaling within the trigeminal system may also contribute to amplification of ipRGC signals. In rodents, bright light activates the trigeminal ganglion and trigeminal nucleus caudalis (72–74). Human studies suggest an interaction of the peripheral trigeminal system and light-mediated pathways as noxious trigeminal stimulation lowers the visual discomfort threshold, and light stimulation lowers trigeminal pain thresholds (11, 75). Studies in the rodent implicate the ipRGCs in this interaction as light aversion following corneal surface damage is attenuated in mice lacking ipRGCs (76). Migraine may induce photophobia through the action of neuropeptides within this trigeminal–thalamic system (77).

We might finally consider the possibility that ipRGC signals for visual discomfort are amplified at the level of the retina. This possibility strikes us as less plausible, given that our results would require a mechanism for selective enhancement of only the class of ipRGC that produces photophobia. Our results also argue against a change in the sensitivity of melanopsin or the cones in migraine under photopic conditions.

There have been varying reports of alteration of cone electroretinogram responses in people with migraine (78, 69), although these studies are also difficult to interpret given possible differences in retinal irradiance between the studied groups (80).

We interpret our results within a modeling approach that assumes that melanopsin and cone signals are integrated within the ipRGCs and that postretinal sites act upon the integrated, log-transformed signal. While this model was not a component of our preregistered experimental protocol, we find that it provides an excellent account of the data. There is abundant evidence in support of the view that melanopsin and cone signals are integrated in the ipRGCs (13, 21–23, 81). We cannot, however, exclude the possibility that cone and melanopsin signals are transmitted from the retina by separate channels and that we are measuring the integration of these signals at some downstream site. Such a postretinal integration is likely to be the case for the percept of brightness—which combines melanopsin and cone contrast—as the postretinal luminance channel originates in signals from the “classical” retinal ganglion cells and must be integrated with signals from melanopsin-containing ipRGCs, perhaps at the level of the lateral geniculate nucleus.

More broadly, there is evidence that expression of melanopsin in eye tissues apart from the retina contributes to photophobia in rodent models (34). Because we placed an artificial pupil between the stimulus and the pharmacologically dilated pupil of the observer, our stimuli illuminated only a small area of the cornea and minimally, the iris. There has also been interest in the contribution of the rods to photophobia in migraine (79), and there is evidence that the rods provide inputs to ipRGCs (82). We sought to minimize the influence of the rods upon our measurements by modulating our stimuli around a photopic background. While there is evidence that rod signals can modulate retinal ganglion cell firing at any light level (83), the amplitude of these effects under photopic conditions is quite small relative to the cones. Further, our prior work indicates that rod

signals do not make a measurable contribution to the pupil response at these background light levels (25).

Conclusion

Our study demonstrates that discomfort from light does not arise as the exclusive action of melanopsin but instead, reflects a signal that integrates cone and melanopsin inputs. The interictal photophobia of migraine is a selective amplification of this integrated signal and one that does not extend to other domains of ipRGC function. We suspect that the enhanced response in migraine to ipRGC signals for discomfort occurs at a postretinal site but cannot yet identify the locus. The modeling approach we adopted here provides a mechanism by which this localization might be pursued, by identifying central sites in which log changes in modeled ipRGC contrast are related to linear modulations of neural activity.

Materials and Methods

We studied 20 participants in each of three groups: MWA, MwoA, and HAF (Table 1). Participants were between 25 and 40 y old and were recruited via digital social media. Headache classification was established using the Penn Online Evaluation of Migraine, which implements the International Classification of Headache Disorders-3-beta criteria (84). Participants with migraine were also required to endorse interictal photophobia (85). Participants completed surveys that assessed behaviors putatively related to ipRGC function (Table 2).

Participants viewed stimuli that targeted specific photoreceptor classes using the technique of silent substitution (86) (Fig. 1C). Each stimulus type was presented at three log-spaced contrast levels: 100, 200, and 400%. These stimuli were produced by a digital light synthesis engine (OneLight Spectra) and tailored for the lens transmittance predicted for the age of each subject. The stimuli were presented through a custom-made eyepiece with a circular, uniform field of 27.5° diameter with the central 5° diameter of the field obscured to minimize macular stimulation. Spectroradiographic measurements were made before and after each session to ensure stimulus quality (SI Appendix, Table S1).

On each of many trials, the participant viewed a pulsed spectral modulation, at one of three contrast levels, designed to target melanopsin, the cones, or both (Fig. 1C). The transition from the background to the stimulation spectrum (melanopsin, cones, or light flux) and the subsequent return

to the background were windowed with a 500-ms half cosine. The total duration of the pulse was 4 s, after which the stimulus field returned to and remained at the background spectrum (Fig. 1D). Twelve seconds after the pulse ended, participants were prompted by an auditory cue to verbally rate their visual discomfort on a 0 to 10 scale. Participants viewed the stimuli through their pharmacologically dilated right eye and a 6-mm-diameter artificial pupil to control retinal irradiance. Infrared video recording of the left eye measured the consensual pupil response during each trial. Each participant viewed at least 12 trials for each crossing of photoreceptor target and contrast, and at least 6 of those trials were required to possess good-quality pupillometry for the subject to be included in the study.

Pupil response was quantified for each trial as the mean percent change in pupil area during the period of 0 to 16 s from stimulus onset, relative to the 0.5 s before stimulus onset. We obtained the median pupil and discomfort response across trials within participant and across participants within groups.

We examined the discomfort and pupil data within a two-stage, nonlinear model (Fig. 3A). The response to a stimulus (either discomfort rating or pupil constriction) is given by

$$Response = m \times \log_{10} \left(\sqrt{\alpha \times C_{Mel}^{\beta} + C_{Cone}^{\beta}} \right) + b,$$

where C_{Mel} and C_{Cone} are the contrasts produced upon the melanopsin and cone photoreceptors by a stimulus, and α , β , m , and b are the four parameters of the model. Nonlinear fitting was performed in MATLAB using `fmincon`, and the variability of parameter estimates within each group was obtained by bootstrap resampling of the data across subjects.

This study was preregistered (SI Appendix, Table S2) and approved by the Institutional Review Board of the University of Pennsylvania. All participants provided informed written consent, and all experiments adhered to the tenets of the Declaration of Helsinki.

Detailed methods are described in SI Appendix, SI Text.

Data Availability. The raw data and analysis code are available through GitHub (<https://github.com/gkaguirrelab/melSquintAnalysis>) (87).

ACKNOWLEDGMENTS. This work was supported by National Eye Institute Grant R01EY024681 (to D.H.B. and G.K.A.) and Core Grant for Vision Research P30 EY001583, National Institute of Neurological Disorders and Stroke Grant R25 NS065745, National Institute on Aging Grant 5T32AG000255-13, and Department of Defense Grant W81XWH-151-0447 (to G.K.A.).

1. K. B. Digre, K. C. Brennan, Shedding light on photophobia. *J. Neuroophthalmol.* **32**, 68–81 (2012).
2. R. Nosedá, R. Burstein, Advances in understanding the mechanisms of migraine-type photophobia. *Curr. Opin. Neurol.* **24**, 197–202 (2011).
3. H. L. Rossi, A. Reuber, Photophobia in primary headaches. *Headache* **55**, 600–604 (2015).
4. G. Selby, J. W. Lance, Observations on 500 cases of migraine and allied vascular headache. *J. Neurol. Neurosurg. Psychiatry* **23**, 23–32 (1960).
5. P. D. Drummond, A quantitative assessment of photophobia in migraine and tension headache. *Headache* **26**, 465–469 (1986).
6. B. K. Rasmussen, R. Jensen, J. Olesen, A population-based analysis of the diagnostic criteria of the International Headache Society. *Cephalalgia* **11**, 129–134 (1991).
7. S. Munjal et al., Most bothersome symptom in persons with migraine: Results from the migraine in America symptoms and treatment (MAST) study. *Headache* **60**, 416–429 (2020).
8. A. Main, A. Dowson, M. Gross, Photophobia and phonophobia in migraineurs between attacks. *Headache* **37**, 492–495 (1997).
9. J. Vanagaite et al., Light-induced discomfort and pain in migraine. *Cephalalgia* **17**, 733–741 (1997).
10. J. V. Vingen, T. Sand, L. J. Stovner, Sensitivity to various stimuli in primary headaches: A questionnaire study. *Headache* **39**, 552–558 (1999).
11. P. A. Kowacs et al., Influence of intense light stimulation on trigeminal and cervical pain perception thresholds. *Cephalalgia* **21**, 184–188 (2001).
12. I. Provencio et al., A novel human opsin in the inner retina. *J. Neurosci.* **20**, 600–605 (2000).
13. D. M. Dacey et al., Melanopsin-expressing ganglion cells in primate retina signal colour and irradiance and project to the LGN. *Nature* **433**, 749–754 (2005).
14. R. Nosedá et al., A neural mechanism for exacerbation of headache by light. *Nat. Neurosci.* **13**, 239–245 (2010).
15. D. M. Berson, Phototransduction by retinal ganglion cells that set the circadian clock. *Science* **295**, 1070–1073 (2002).
16. K. Thapan, J. Arendt, D. J. Skene, An action spectrum for melatonin suppression: Evidence for a novel non-rod, non-cone photoreceptor system in humans. *J. Physiol.* **535**, 261–267 (2001).
17. R. J. Lucas et al., Diminished pupillary light reflex at high irradiances in melanopsin-knockout mice. *Science* **299**, 245–247 (2003).
18. R. J. Lucas, R. H. Douglas, R. G. Foster, Characterization of an ocular photopigment capable of driving pupillary constriction in mice. *Nat. Neurosci.* **4**, 621–626 (2001).
19. P. D. R. Gamlin et al., Human and macaque pupil responses driven by melanopsin-containing retinal ganglion cells. *Vision Res.* **47**, 946–954 (2007).
20. J. M. Stringham, K. Fuld, A. J. Wenzel, Action spectrum for photophobia. *J. Opt. Soc. Am. A Opt. Image Sci. Vis.* **20**, 1852–1858 (2003).
21. J. A. Perez-Leon, E. J. Warren, C. N. Allen, D. W. Robinson, R. L. Brown, Synaptic inputs to retinal ganglion cells that set the circadian clock. *Eur. J. Neurosci.* **24**, 1117–1123 (2006).
22. K. Y. Wong, F. A. Dunn, D. M. Graham, D. M. Berson, Synaptic influences on rat ganglion-cell photoreceptors. *J. Physiol.* **582**, 279–296 (2007).
23. S. Weng, M. E. Estevez, D. M. Berson, Mouse ganglion-cell photoreceptors are driven by the most sensitive rod pathway and by both types of cones. *PLoS One* **8**, e66480 (2013).
24. M. Spitschan, S. Jain, D. H. Brainard, G. K. Aguirre, Opponent melanopsin and S-cone signals in the human pupillary light response. *Proc. Natl. Acad. Sci. U.S.A.* **111**, 15568–15572 (2014).
25. H. McAdams, A. Igdalova, M. Spitschan, D. H. Brainard, G. K. Aguirre, Pulses of melanopsin-directed contrast produce highly reproducible pupil responses that are insensitive to a change in background radiance. *Invest. Ophthalmol. Vis. Sci.* **59**, 5615–5626 (2018).
26. R. B. Lipton, M. E. Bigal, Migraine: Epidemiology, impact, and risk factors for progression. *Headache* **45** (suppl. 1), S3–S13 (2005).
27. J. Olesen, T. J. Steiner, The international classification of headache disorders, 2nd edn (ICDH-II). *Cephalalgia* **24** (suppl. 1), 9–160 (2004).
28. W. F. Stewart, R. B. Lipton, A. J. Dowson, J. Sawyer, Development and testing of the Migraine Disability Assessment (MIDAS) questionnaire to assess headache-related disability. *Neurology* **56** (6, suppl. 1), S20–S28 (2001).
29. M. Yang, R. Rendas-Baum, S. F. Varon, M. Kosinski, Validation of the Headache Impact Test (HIT-6™) across episodic and chronic migraine. *Cephalalgia* **31**, 357–367 (2011).
30. E. G. Conlon, W. J. Lovegrove, E. Chekaluk, P. E. Pattison, Measuring visual discomfort. *Vis. Cogn.* **6**, 637–663 (1999).

31. L. Bossini *et al.*, Sensibilità alla luce e psicopatologia: Validazione del Questionario per la Valutazione della Fotosensibilità (Q.V.F.). *Med. Psicosom.* **51**, 167–176 (2006).
32. J. A. Horne, O. Ostberg, A self-assessment questionnaire to determine morningness-eveningness in human circadian rhythms. *Int. J. Chronobiol.* **4**, 97–110 (1976).
33. N. E. Rosenthal, G. H. Bradt, T. A. Wehr, *Seasonal Pattern Assessment Questionnaire*, (National Institute of Mental Health, Washington DC, 1987).
34. A. Matynia *et al.*, Peripheral sensory neurons expressing melanopsin respond to light. *Front. Neural Circuits* **10**, 60 (2016).
35. R. N. Shepard, Toward a universal law of generalization for psychological science. *Science* **237**, 1317–1323 (1987).
36. R. F. Quick Jr., A vector-magnitude model of contrast detection. *Kybernetik* **16**, 65–67 (1974).
37. A. J. Zele, P. Adhikari, D. Cao, B. Feigl, Melanopsin and cone photoreceptor inputs to the afferent pupil light response. *Front. Neural.* **10**, 529 (2019).
38. S. B. Baver, G. E. Pickard, P. J. Sollars, G. E. Pickard, Two types of melanopsin retinal ganglion cell differentially innervate the hypothalamic suprachiasmatic nucleus and the olivary pretectal nucleus. *Eur. J. Neurosci.* **27**, 1763–1770 (2008).
39. S. Hattar, H. W. Liao, M. Takao, D. M. Berson, K. W. Yau, Melanopsin-containing retinal ganglion cells: Architecture, projections, and intrinsic photosensitivity. *Science* **295**, 1065–1070 (2002).
40. X. Zhao, B. K. Stafford, A. L. Godin, W. M. King, K. Y. Wong, Photoresponse diversity among the five types of intrinsically photosensitive retinal ganglion cells. *J. Physiol.* **592**, 1619–1636 (2014).
41. U. Grünert, P. R. Jusuf, S. C. S. Lee, D. T. Nguyen, Bipolar input to melanopsin containing ganglion cells in primate retina. *Vis. Neurosci.* **28**, 39–50 (2011).
42. S. Neumann, S. Haverkamp, O. N. Aufferkorte, Intrinsically photosensitive ganglion cells of the primate retina express distinct combinations of inhibitory neurotransmitter receptors. *Neuroscience* **199**, 24–31 (2011).
43. P. R. Jusuf, S. C. S. Lee, J. Hannibal, U. Grünert, Characterization and synaptic connectivity of melanopsin-containing ganglion cells in the primate retina. *Eur. J. Neurosci.* **26**, 2906–2921 (2007).
44. J. Hannibal, A. T. Christiansen, S. Heegaard, J. Fahrenkrug, J. F. Kiilgaard, Melanopsin expressing human retinal ganglion cells: Subtypes, distribution, and intraretinal connectivity. *J. Comp. Neurol.* **525**, 1934–1961 (2017).
45. J. Hannibal *et al.*, Melanopsin is expressed in PACAP-containing retinal ganglion cells of the human retinohypothalamic tract. *Invest. Ophthalmol. Vis. Sci.* **45**, 4202–4209 (2004).
46. S. K. Chen, T. C. Badea, S. Hattar, Photoentrainment and pupillary light reflex are mediated by distinct populations of ipRGCs. *Nature* **476**, 92–95 (2011).
47. P. A. Barrionuevo *et al.*, Assessing rod, cone, and melanopsin contributions to human pupil flicker responses. *Invest. Ophthalmol. Vis. Sci.* **55**, 719–727 (2014).
48. P. A. Barrionuevo, D. Cao, Luminance and chromatic signals interact differently with melanopsin activation to control the pupil light response. *J. Vis.* **16**, 29 (2016).
49. M. Spitschan *et al.*, The human visual cortex response to melanopsin-directed stimulation is accompanied by a distinct perceptual experience. *Proc. Natl. Acad. Sci. U.S.A.* **114**, 12291–12296 (2017).
50. T. M. Brown *et al.*, Melanopsin-based brightness discrimination in mice and humans. *Curr. Biol.* **22**, 1134–1141 (2012).
51. M. Yamakawa, S. I. Tsujimura, K. Okajima, A quantitative analysis of the contribution of melanopsin to brightness perception. *Sci. Rep.* **9**, 7568 (2019).
52. T. DeLaywer, S. I. Tsujimura, K. Shinomori, Relative contributions of melanopsin to brightness discrimination when hue and luminance also vary. *J. Opt. Soc. Am. A Opt. Image Sci. Vis.* **37**, A81–A88 (2020).
53. A. J. Zele, P. Adhikari, B. Feigl, D. Cao, Cone and melanopsin contributions to human brightness estimation. *J. Opt. Soc. Am. A Opt. Image Sci. Vis.* **35**, B19–B25 (2018).
54. R. Storchi *et al.*, Melanopsin-driven increases in maintained activity enhance thalamic visual response reliability across a simulated dawn. *Proc. Natl. Acad. Sci. U.S.A.* **112**, E5734–E5743 (2015).
55. M. Hatori *et al.*, Inducible ablation of melanopsin-expressing retinal ganglion cells reveals their central role in non-image forming visual responses. *PLoS One* **3**, e2451 (2008).
56. R. J. Lucas, Diminished pupillary light reflex at high irradiances in melanopsin-knockout mice. *Science* **299**, 245–247 (2003).
57. M. Cambron, H. Maertens, K. Paemeleire, L. Crevits, Autonomic function in migraine patients: Ictal and interictal pupillometry. *Headache* **54**, 655–662 (2014).
58. D. E. Harle, J. S. Wolffsohn, B. J. W. Evans, The pupillary light reflex in migraine. *Ophthalmic Physiol. Opt.* **25**, 240–245 (2005).
59. O. E. Eren, R. Ruscheweyh, C. Schankin, F. Schöberl, A. Straube, The cold pressor test in interictal migraine patients—different parasympathetic pupillary response indicates dysbalance of the cranial autonomic nervous system. *BMC Neurol.* **18**, 41 (2018).
60. P. D. Drummond, Pupil diameter in migraine and tension headache. *J. Neurol. Neurosurg. Psychiatry* **50**, 228–230 (1987).
61. M. M. Cortez, N. A. Rea, L. A. Hunter, K. B. Digre, K. C. Brennan, Altered pupillary light response scales with disease severity in migrainous photophobia. *Cephalalgia* **37**, 801–811 (2017).
62. M. M. Cortez, N. Rae, L. Millsap, N. McKean, K. C. Brennan, Pupil cycle time distinguishes migraineurs from subjects without headache. *Front. Neural.* **10**, 478 (2019).
63. K. A. Roeklein *et al.*, Melanopsin gene variations interact with season to predict sleep onset and chronotype. *Chronobiol. Int.* **29**, 1036–1047 (2012).
64. K. A. Roeklein *et al.*, A missense variant (P10L) of the melanopsin (OPN4) gene in seasonal affective disorder. *J. Affect. Disord.* **114**, 279–285 (2009).
65. K. A. Roeklein *et al.*, Melanopsin, photosensitive ganglion cells, and seasonal affective disorder. *Neurosci. Biobehav. Rev.* **37**, 229–239 (2013).
66. J. Johnson *et al.*, Melanopsin-dependent light avoidance in neonatal mice. *Proc. Natl. Acad. Sci. U.S.A.* **107**, 17374–17378 (2010).
67. A. Routtenberg, M. Strop, J. Jerdan, Response of the infant rat to light prior to eyelid opening: Mediation by the superior colliculus. *Dev. Psychobiol.* **11**, 469–478 (1978).
68. D. S. McNeill *et al.*, Development of melanopsin-based irradiance detecting circuitry. *Neural Dev.* **6**, 8 (2011).
69. P. J. Goadsby *et al.*, Pathophysiology of migraine: A disorder of sensory processing. *Physiol. Rev.* **97**, 553–622 (2017).
70. W. M. Mulleners, E. P. Chronicle, J. E. Palmer, P. J. Koehler, J. W. Vredeveld, Visual cortex excitability in migraine with and without aura. *Headache* **41**, 565–572 (2001).
71. D. J. Hodkinson *et al.*, Increased amplitude of thalamocortical low-frequency oscillations in patients with migraine. *J. Neurosci.* **36**, 8026–8036 (2016).
72. V. Marek *et al.*, Implication of melanopsin and trigeminal neural pathways in blue light photosensitivity *in vivo*. *Front. Neurosci.* **13**, 497 (2019).
73. K. Okamoto, R. Thompson, A. Tashiro, Z. Chang, D. A. Bereiter, Bright light produces Fos-positive neurons in caudal trigeminal brainstem. *Neuroscience* **160**, 858–864 (2009).
74. K. Okamoto, A. Tashiro, Z. Chang, D. A. Bereiter, Bright light activates a trigeminal nociceptive pathway. *Pain* **149**, 235–242 (2010).
75. P. D. Drummond, A. Woodhouse, Painful stimulation of the forehead increases photophobia in migraine sufferers. *Cephalalgia* **13**, 321–324 (1993).
76. A. Matynia *et al.*, Light aversion and corneal mechanical sensitivity are altered by intrinsically photosensitive retinal ganglion cells in a mouse model of corneal surface damage. *Exp. Eye Res.* **137**, 57–62 (2015).
77. E. A. Kaiser, A. F. Russo, CGRP and migraine: Could PACAP play a role too? *Neuropeptides* **47**, 451–461 (2013).
78. R. Nosedá *et al.*, Migraine photophobia originating in cone-driven retinal pathways. *Brain* **139**, 1971–1986 (2016).
79. C. A. Bernstein *et al.*, The migraine eye: Distinct rod-driven retinal pathways' response to dim light challenges the visual cortex hyperexcitability theory. *Pain* **160**, 569–578 (2019).
80. O. A. Mahroo, Pupil area and photopigment spectral sensitivity are relevant to study of migraine photophobia. *Brain* **140**, e2 (2017).
81. K. Y. Wong, A retinal ganglion cell that can signal irradiance continuously for 10 hours. *J. Neurosci.* **32**, 11478–11485 (2012).
82. S. K. Lee, T. Sonoda, T. M. Schmidt, M1 intrinsically photosensitive retinal ganglion cells integrate rod and melanopsin inputs to signal in low light. *Cell Rep.* **29**, 3349–3355.e2 (2019).
83. A. Tikidji-Hamburyan *et al.*, Rods progressively escape saturation to drive visual responses in daylight conditions. *Nat. Commun.* **8**, 1813 (2017).
84. E. A. Kaiser, A. Igdalova, G. K. Aguirre, B. Cucchiara, A web-based, branching logic questionnaire for the automated classification of migraine. *Cephalalgia* **39**, 1257–1266 (2019).
85. J. Y. Choi *et al.*, Usefulness of a photophobia questionnaire in patients with migraine. *Cephalalgia* **29**, 953–959 (2009).
86. O. Estévez, H. Spekreijse, The “silent substitution” method in visual research. *Vision Res.* **22**, 681–691 (1982).
87. H. McAdams, G. Frazzetta, G. Aguirre, Analysis code for pupil tracking data for the OLApproach_Squint project. GitHub. <https://github.com/gkaguirrelab/melSquintAnalysis>. Deposited 3 June 2020.